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REVIEW

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Quality of fresh and fresh-cut produce impacted by nonthermal physical technologies intended to enhance microbial safety

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ABSTRACT

Nonthermal physical intervention technologies are able to reduce populations of foodborne pathogens on/in fresh produce. As highly perishable and living organisms, fresh produce is inherently sensitive to any physical or chemical treatment in terms of quality damage. The aims of the present review are to summarize current knowledge on non-thermal technologies (ionizing radiation, UV and pulsed light, ultrasound, high hydrostatic pressure, and cold plasma) with an emphasis on their impact on quality of fresh produce and to discuss advantages, disadvantages, and considerations for the commercialization of each technology. The impact of nonthermal physical technologies on fresh produce quality is related to pathogen inactivation mechanisms of each individual technology, and the nature and intensity of changes in quality due to the technologies depend on the treatment intensity/time and other processing conditions. Common symptoms of quality deterioration due to nonthermal processing include tissue softening, browning, and loss of nutrients. In general, there is a lack of systematic assessment, particularly sensory evaluations using taste panels of the product quality after treatments. For emerging technologies, such as cold plasma, more studies are necessary in order to assess quality changes during post-treatment storage at relevant temperatures. Quality of fresh produce must be carefully investigated to facilitate the commercialization of technologies.

KEYWORDS

Ionizing radiation; ultraviolet; pulsed light; ultrasound; high pressure; fresh-cut

Introduction

Fresh fruits and vegetables are rich in carbohydrates, fibers, vitamins, and other nutrients such as antioxidants and have been highly recommended by health professionals and organizations as part of a well-balanced daily diet to reduce the risk of heart disease, stroke, and development of certain types of cancer, diabetes, and many other health issues (Lavin and Lloyd [2012](#page-18-0); Baselice et al. [2017;](#page-16-0) Singla, Chaturvedi, and Sandhu [2020](#page-20-0)). In order to meet the changes in lifestyle and requirements of convenience, fresh produce is minimally processed to 'fresh-cut' products. Fresh-cut produce, refers to 'any fresh fruit or vegetable or combination thereof that has been physically altered from its whole state after being harvested from the field (e.g., by chopping, dicing, peeling, ricing, shredding, slicing, spiralizing, or tearing) without additional processing (such as blanching or cooking)' (USFDA [2019\)](#page-20-0). With rising income, increasingly busy lifestyles, and demands for convenience and healthbenefits, the sale of fresh-cut produce has increased substantially over the last 2–3 decades in many countries (Del Gobbo et al. [2015](#page-16-0); Baselice et al. [2017](#page-16-0)).

Unfortunately, the number of outbreaks of foodborne illnesses associated with fresh and fresh-cut produce has increased concomitantly in the U.S. and many other coun-tries in recent years (Callejón et al. [2015](#page-16-0); Del Gobbo et al. [2015](#page-16-0); Bintsis [2017](#page-16-0)). The U.S. Centers for Disease Control and Prevention (CDC) estimated the number of foodborne illness from 31 major pathogens (Scallan et al. [2011\)](#page-20-0). The 31 known pathogens acquired in the United States caused 9.4 million episodes of foodborne illness, 55,961 hospitalizations, and 1351 deaths every year. While norovirus has been shown to be responsible for the majority of produce-related illnesses in the U.S. and Europe, bacterial pathogens such as Salmonella enterica, Shiga toxin producing Escherichia coli, and Listeria monocytogenes often cause more serious harm and complications and number of deaths (Callejón et al. [2015](#page-16-0); Carstens, Salazar, and Darkoh [2019\)](#page-16-0). In addition, foodborne illnesses result in significant economic losses. According to the United States Department of Agriculture (USDA), the cost of foodborne illness is estimated to be \$10–83 billion per year in the U.S. (USDA, 2014). The total number of reported outbreaks of foodborne illnesses associated with all foods decreased by 38% during 1998–2013 in the U.S. (Bennett et al. [2018\)](#page-15-0). The percentage of outbreaks

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attributed to fresh produce among all outbreaks, however, increased from 8% during 1998–2001 to 16% during 2010–2013, suggesting that many other food industry sectors have improved their practices, while the fresh produce industry needs to adopt further preventive and control measures. The fresh produce industry is in urgent need of affective intervention technologies. Postharvest intervention is of vital importance to enhance the microbial safety of fresh produce. Over the years, many different types of postharvest intervention technologies and treatments have been developed including chemical, physical, and biological means. Some of the technologies show promise in inactivating human pathogens on fresh produce. However, there are few technologies that are commercialized. There are many reasons for the lack of adopting technology. One of reasons may be due to their undesirable effects on quality of fresh and fresh-cut produce, including surface discoloration), softening, and loss of nutrients such as vitamin C. While safety is a prerequisite of any food product, product quality and shelf-life often determine consumer acceptance and the success of any new processing technology. Quality of fresh produce is determined by a combination of parameters including appearance, texture, flavor, and nutritional value (Kader [2002;](#page-17-0) Barrett, Beaulieu, and Shewfelt [2010](#page-16-0)).

There are many recent reviews on various interventions, including conventional and emerging technologies in order to improve microbial safety of fresh and fresh-cut fruits and vegetables (Gil, Allende, and Selma [2011;](#page-17-0) Alexandre, Brandão, and Silva [2012;](#page-15-0) Pasha et al. [2014;](#page-19-0) Smetanska, Hunaefi, and Barbosa-Cánovas [2013;](#page-20-0) Warriner and Namvar [2013;](#page-21-0) Artes and Allende [2014](#page-15-0); Mahajan et al. [2014;](#page-18-0) Nicola and Fontana [2014;](#page-19-0) Gil et al. [2015](#page-17-0); Ali et al. [2018](#page-15-0); Erkan and Yıldırım [2017](#page-16-0); Ma et al. [2017](#page-18-0); Mahajan et al. [2017;](#page-18-0) Singh, Walia, and Batra [2018;](#page-20-0) De Corato [2020](#page-16-0)). The present review focuses on the impact of physical interventions on quality of fresh and fresh-cut produce. We hope that this review can be useful for future research directions to minimize the unwanted quality changes, due to intervention technologies, and ultimately aiding to the implantation of effective technologies to enhance microbial safety of fresh produce. Non-thermal physical interventions to be discussed include ionizing radiation, UV, pulsed light, ultrasound, high hydrostatic pressure processing, and cold plasma.

Ionizing radiation

There are three common types of ionizing radiation: gamma ray, electron beam, and X-ray (Fan and Niemira [2020](#page-16-0)). Gamma rays are produced from a nuclear source (commonly cobalt-60 or cesium-137) and have excellent penetration capability. Electron beams produced often by accelerators, have low penetration abilities, but permits fast processing speeds (in seconds). X-rays have high penetration abilities and are converted from electron beams. All three types of ionizing radiation, at similar doses, have similar effects on human pathogens and product quality because all three exerts its effects through two mechanisms: direct effect and indirect effect (Fan [2012](#page-16-0)). For the direct effect, ionizing radiation directly causes damages to cell components such as DNA, carbohydrates, and lipids. For the indirect effect, free radicals and reactive species (such as hydrated electrons, hydrogen atoms, and hydroxyl radicals) from radiolysis of water react with cells or food components (Simic [1983](#page-20-0)). In fresh produce, water is the major component, therefore, the indirect effect is the major mechanism for the changes induced by ionizing irradiation.

At low doses $(\leq 1 \text{ kGy})$, irradiation reduces populations of human pathogenic bacteria and spoilage microorganisms without significant losses in quality parameters such as color, firmness or sensory attributes of many fresh produce items, for example spinach and tomato (Fan, Niemira, and Sokorai 2003; Fan and Sokorai [2008\)](#page-17-0) ([Table 1\)](#page-3-0). However, 2 kGy gamma irradiation decreased vitamin C content by 50% in fresh cilantro and 3 kGy had higher decay rate and offodor scores due to irradiation-induced damages after 14 days of storage at 3°C (Fan, Niemira, and Sokorai [2003a\)](#page-17-0). Similarly, 2 kGy irradiation treatment reduced ascorbic acid content of spinach by 75% after 14 days storage at 4° C while the appearance, aroma, texture, flavor, and overall ratings were not affected (Fan and Sokorai [2011](#page-17-0)). Another study showed that 1.0 kGy irradiation alleviated the loss in vitamin C content and inhibited polyphenol oxidase activity of fresh-cut lettuce during storage (Zhang et al. [2006](#page-21-0)). When applied as a disinfection technology, X-ray irradiation at doses up to 0.8 kGy did not affect soluble solids content, titratable acidity, or fructose concentrations of dragon fruit. However, glucose, sucrose, and total sugar concentrations decreased linearly with increasing doses (Wall and Khan [2008\)](#page-21-0). Electron beam (1.0, 1.5 and 3.2 kGy) resulted in a loss in firmness of romaine lettuce (Han et al. [2004\)](#page-17-0) and 3.2 kGy led to the loss of firmness and aroma of blueberry fruit (Moreno et al. [2007\)](#page-19-0). It has been shown that low-dose electron beam treatments (1.0–1.5 kGy) increased carotene content in cantaloupes (Castell-Perez et al. [2004](#page-16-0)), while 1 kGy electron beam irradiation did not cause any effect on firmness, color, or flavor of fresh-cut watermelon (Smith et al. [2017\)](#page-20-0).

There are some studies demonstrating that some fresh produce items can tolerate higher doses and even showing beneficial effects during post-irradiation storage. For example, irradiation (2 kGy) inhibited the change of titratable acidity and pH and reduced the population of microorganisms in Chinese cabbage during refrigerated storage (Ahn et al. [2005\)](#page-15-0). Radiation (2.5 kGy) resulted in a lower spoilage rate and maintained ultrastructure of blueberries after 21–35 days of storage, compared to the nontreated control (Wang and Meng [2016\)](#page-21-0). Irradiation (2 kGy) improved microbial safety without significant loss on total carotenes, ascorbic acid, and sucrose content of carrots (Kamat et al. [2005\)](#page-18-0). It has also been shown that irradiation (2 kGy) reduced browning of cabbage without causing changes in phenolic acid content, even though a 1.4–fold decrease in phenylalanine ammonia lyase activity was observed (Banerjee et al. [2015\)](#page-15-0) ([Table 1](#page-3-0)). A 5 kGy radiation preserved antioxidant activity and total flavonoid content and increased content of total phenolics of watercress (Pinela

Table 1. Effects of ionizing radiation on quality attributes of fresh and fresh-cut produce.

Treatment conditions (concentration,	
time, intensity, dose etc.)	Tvp

et al. [2016](#page-20-0)). However, the high dose likely resulted in the deterioration of sensory quality.

Since irradiation can result in quality deterioration, research has been conducted to combine ionizing radiation with other treatments to minimize the losses. For example, pre-irradiation warm water dipping combined with modified atmosphere packaging reduced undesirable effects of irradiation on quality of fresh-cut iceberg lettuce including firmness, vitamin C, and total antioxidants content (Fan, Toivonen, et al. [2003](#page-17-0)); calcium ascorbate treatment could alleviate the firmness loss and browning caused by ionizing radiation on fresh-cut 'Gala' apples (Fan et al. [2005](#page-17-0)). Combination of carboxymethyl cellulose coating and gamma irradiation resulted in chlorophyll retention and a lower rot rate for plums and peaches (Hussain et al. [2015,](#page-17-0) [2016](#page-17-0)).

[Table 1](#page-3-0) lists the studies related to ionizing radiation effects on quality of fresh and fresh-cut produce. Overall, ionizing radiation at low doses (1 kGy) did not significantly affect sensory quality or nutrient quality of fresh produce. At low doses, more than 5 log reductions of human pathogens on fresh produce could be achieved (Niemira [2008](#page-19-0)). Higher doses may result in the loss of nutrients, particularly vitamin C content. Irradiation may also increase phenolic contents and other phytochemicals in some produce items. The effect of irradiation depends on dose, type of fresh produce, and storage conditions. Another major effect of irradiation on fresh produce quality is immediate loss of firmness after irradiation. The loss of firmness may be related to irradiation-induced changes in cell wall components, such as hydrolysis of pectin, cell wall modifications, and pectin methyl esterase activity (Melo et al. [2018\)](#page-18-0).

Even though low dose irradiation has been shown to be effective in reducing human pathogens on fresh produce without significant effects on produce quality, its application is very limited. The current uses of irradiating are mainly for phytosanitary purposes in order to meet quarantine requirements of disinfestation (Roberts and Follett [2017](#page-20-0)). One of the major factors limiting the commercial application of ionizing radiation is reluctance in consumer acceptance of the technology, mostly due to the misinformation and concern on using radioactive materials (Bearth et al. 2019). Educating consumers about the benefits and nature of ionizing radiation will aid in the acceptance and application of the technology. X-ray and electron beam technologies which do not involve the use of radioactive isotopes, may have advantages in terms of consumer acceptance. Overall, ionizing radiation is a relatively mature non-thermal technology and can be applied for niche products and markets to enhance microbial safety of foods, inhibit sprouting of tubers, and disinfesting against insects.

UV and pulsed light

Basing on wavelength, ultraviolet as a non-ionizing radiation can be divided into three groups: 100-280 nm (UV-C), 280–315 nm (UV-B), and 315–400 nm (UV-A). UV-C is the most effective germicidal UV light. The germicidal effect of UV-C light is a result of its ability to damage DNA or RNA of a microorganism and consequently blocking cell replica-tion (Sinha and Häder [2002;](#page-20-0) Escalona et al. [2010\)](#page-16-0). UV-C induces a variety of mutagenic and cytotoxic DNA lesions via photocatalytic formation of cyclobutane thymine dimers. Therefore UV-C light can effectively inactivate microorganisms, including pathogenic and spoilage microorganisms. UV-C technology does not leave chemical residue, nor requires extensive worker-protection equipment (Yousef and

Marth [1988\)](#page-21-0). Application of UV technology in continuous and pulse modes for processing whole and fresh-cut fruits have been discussed (Koutchma, Orlowska, and Zhu [2018\)](#page-18-0). UV light treatment of fresh fruits and vegetable surfaces have been investigated to decontaminate surfaces, reduce decay, and enhance shelf life and quality (Turtoi [2013\)](#page-20-0) ([Table 2\)](#page-5-0).

UV-C has been found to reduce microbial populations on fresh and fresh-cut produce without significant effects on quality. For example, lower dose UV-C light $(4.1 \text{ kJ} \cdot \text{m}^{-2}$ at 254 nm) can achieve microbial reductions comparable to aqueous sanitizers, such as chlorine and ozone without affecting juice leakage, color, and overall visual quality of fresh-cut watermelon (Fonseca et al. 2006). On the surface of button mushroom, UV-C $(0.45-3.15 \text{ kJ} \bullet \text{ m}^{-2})$ resulted in 0.67-1.13 log (CFU \bullet g⁻¹) reductions of *E. coli* O157:H7 and 0.63–0.89 log (CFU \bullet g⁻¹) total aerobic plate, without deterioration effects on quality (Guan, Fan, and Yan [2012\)](#page-17-0). Furthermore, the UV-C treatment inhibited surface lesion and extended shelf-life, presumably due to its antimicrobial effect on Pseudomonas spp. that caused the lesion development. Water-assisted UV (samples were treated by UV while being immersed in agitated water) was found very effective in inactivating Salmonella on tomatoes and fresh-cut lettuce (Guo, Huang, and Chen [2017\)](#page-17-0). In addition, combination of $2 \text{ kJ} \cdot \text{m}^{-2}$ UV-C and active modified atmosphere conditions, applied to cherry tomatoes stored at low temperatures, delayed S. Typhimurium growth, improved color and firmness, and lycopene content (Choi et al. [2015\)](#page-16-0).

UV-C may be able to reduce browning of fresh and fresh-cut produce. UV-C treatment (75 W for 5 and 10 min) controlled browning of fresh-cut lotus root by inactivation of polyphenol oxidase, peroxidase, and phenylalanine ammonia lyase (enzymes involved in browning) and reduced soluble quinone content without causing changes in soluble solids content or firmness (Wang et al. [2019](#page-21-0)). Similarly, UV-C combined with warm water washing $(45\degree C, 120\degree s)$ deactivated the phenylalanine ammonia lyase, polyphenol oxidase, and peroxidase, as well as improved sensory qual-ities of fresh-cut endive (Hägele et al. [2016](#page-17-0)). Other ranges of UV may also have a positive role in inhibiting browning of apples and pears (Lante, Tinello, and Nicoletto [2016\)](#page-18-0). However, UV-C is also found accelerating initial browning in cut-apples (Gómez et al. [2010\)](#page-17-0) and mushrooms (Guan, Fan, and Yan [2012](#page-17-0)). Although samples treated with UV-C had more severe browning immediately after treatment, the control mushrooms browned faster during storage than the UV-C treated samples (Guan, Fan, and Yan [2012\)](#page-17-0). The effect of UV may depend on dose, type of fresh produce, treatment time, and storage.

UV-C treatment could reduce decay in fruits and vegetables such as grapefruit (D'hallewin et al. [2000\)](#page-16-0). Exposure of fresh-cut melon to UV-C light led to 2 log reductions of both total viable count and Enterobacteriaceae. Low microbial populations were maintained during storage with no significant effects on product color or firmness (Manzocco, Da Pieve, and Maifreni [2011](#page-18-0)). In addition, all three types of UV are able to induce resistance of fruits and vegetables to

Table 2. Effects of UV-C on quality attributes and microbial populations of fresh and fresh-cut produce.

Treatment conditions (concentration, time, intensity, dose etc.)	Type of fresh produce	Effect on quality	Reference
UV-C	fresh-cut watermelon	reduced microbial populations	Fonseca et al. 2006
4.1 $\text{kl} \cdot \text{m}^{-2}$ at 254 nm		without affecting juice leakage, color, and	
UV-C	fresh-cut lotus root	overall visual quality controlled browning by inactivation of	Wang et al. 2019
0, 0.3, 1.5, 3, 6, 12 $\text{kl} \cdot \text{m}^{-2}$		polyphenol oxidase, peroxidase, and	
corresponding to 0, 1, 5, 10, 20,		phenylalanine ammonia lyase and reduce	
and 40 min at 4° C respectively		soluble quinone content without change on	
		soluble solids content and firmness (5	
UV-C	fresh-cut endive	and 10 min) disactivated the phenylalanine ammonia-lyase,	Hägele et al. 2016
1.2 $\text{kl} \cdot \text{m}^{-2}$ and warm (45 °C, 120 s)		polyphenol oxidase, and peroxidase and	
water washing		improved sensory qualities significantly	
Water-assisted UV 29 mW•cm ⁻² for	grape tomato and fresh-cut lettuce	Reduced populations of Salmonella by 3.84 and	Guo et al. 2017
2min (samples were treated by UV		1.79 log $CFU \bullet g^{-1}$ from 6.14 and 8.23 log $CFU \bullet g^{-1}$, on tomato and lettuce, respectively.	
while being immersed in agitated water)			
UV-C	grape	$0.5 \text{ kJ} \cdot \text{m}^{-2}$ reduce decay, irradiation at dosages	D'hallewin et al. 2000
0.5, 1.5, or 3.0 $\text{kJ}\cdot\text{m}^{-2}$		>0.5 kJ·m ⁻² did not further improve decay	
		control and caused rind browning and	
UV-C	pepper	necrotic peel reduced decay and chilling injury	Vicente et al. 2005
7 k Jom $^{-2}$			
UV-C	cut-apple	accelerated browning	Gómez et al. 2010
10, 15 or 25 min (5.6 ± 0.3)			
8.4 ± 0.5 and 14.1 ± 0.9 kJ \bullet m ⁻² ,			
respectively) UV-C	peeled garlic	increased antioxidant content including total	Park and Kim 2015
$2 \text{ kJ} \cdot \text{m}^{-2}$		polyphenol, flavonoid, apigenin and	
		quercetin levels, improved firmness during	
UV-C	button mushroom	storage for peeled garlic after storage	Guan et al. 2012
$0.45 - 3.15$ kJom ⁻²		0.67-1.13 and 0.63-0.89 log CFU e^{-1} reduction of E. coli O157:H7 and total aerobic plate,	
		respectively, inhibited surface lesion and	
		extended shelf-life without deterioration	
		effects on quality	
UV-C 0, 3.7×10^3 Jom ⁻² ,	tomato	a reduction in respiration rate, ethylene production, and an increase in putrescine	Maharaj et al. 1999
24.4×10^3 Jom ⁻²			
UV-C	watermelon	increased total antioxidant capacity with slight	Artés-Hernández et al. 2010
1.6, 2.8, 4.8 and 7.2 $\text{kl} \cdot \text{m}^{-2}$		effect on color, lycopene	
UV-C 0, 1200, 6000 and 12,000 $\text{J} \cdot \text{m}^{-2}$	fresh-cut melon	inhibited microbial growth, decreased leakage, and induced better flavor	Manzocco et al. 2011
$UV-C$ 2 kJ \bullet m ⁻² and passive and active	cherry tomato	improved the color and firmness, and lycopene	Choi et al. 2015
modified atmosphere		content and delayed S. typhimurium growth	
UV-C 0, 3.4, 7.2 and 10.5 $\text{kl} \cdot \text{m}^{-2}$	Tahitian limes	the highest UV-C treatment (10.5 $\text{kl} \cdot \text{m}^{-2}$)	Pristijono et al. 2019
		maintained low ethylene production and low respiration rates and had a 60%	
		acceptability index after 28 days storage	
		with no differences between the different	
		UV-C intensities and no effect on fruit	
UV-B	sweet basil	weight loss, TSS or TA contents 3.60 $W/m2$ was the best which increased total	Ghasemzadeh et al. 2016
2.30, 3.60, and 4.80 Wom ⁻² for 4,		flavonoid, total phenol, quercetin, catechin,	
6, 8, and 10 h respectively		kaempferol, rutin, ferulic acid, gallic acid	
		and chalcone synthase (CHS) activity,	
UV-A	apple and pear	induced cinnamic acid and luteolin antibrowning effects on cut surfaces	Lante et al. 2016
2.43×10^{-3} Wom ⁻² for up to			
60 min			
UV-C	tomato	enhanced antioxidant capacity by increasing	Pataro et al. 2015
1, 2, 4, 8 Jocm ⁻² (0.95, 1.9, 3.8 and 7.6 s each side, respectively)		lycopene, total carotenoid, phenolic compounds content without effect on pH	
		and soluble solid content	
UV-C	spinach	In stage II (baby spinach), a decrease in	Martínez-Sánchez et al. 2019
1.5, 3 Jocm ⁻²		vitamin C content but an increase with total	
		plate count along with the number of	
		applications was observed after applying 3 J •cm ⁻² . Maintained the meshophilic and	
		yeast counts during the spinach	
		development in the field and reduced the	
UV-C		mesophilic count after storage. Treatment decreased the malate content and no	Onik et al. 2020
9 Jocm $^{-2}$	Apple	changes in sugar content was found, higher	
		ratio of total sugars to total organic acids.	

postharvest spoilage by activating genes encoding pathogenesis-related proteins, leading to the accumulations of phytoalexins.

It is well known that UV-C increased antioxidants of fresh and fresh-cut produce by promoting synthesis of secondary metabolic compounds. For example, UV-C $(2 \text{ kJ} \cdot \text{m}^{-2})$ increased antioxidants content including total polyphenol, flavonoid, apigenin, and quercetin levels of peeled garlic during storage (Park and Kim [2015](#page-19-0)). In another study, UV-C increased total antioxidant capacity with slight effects on lycopene of fresh-cut watermelon (Artés-Hernández et al. [2010\)](#page-15-0). In addition, research has shown that UV-B (3.60 $W \bullet m^{-2}$ for up to 10 h) increased the amount of total flavonoid, total phenol, and several individual phenolic compounds of sweet basil, which resulted in higher antioxidant capacity and pharmaceutical properties (Ghasemzadeh et al. [2016](#page-17-0)). Furthermore, UV-C enhanced antioxidant capacity of tomato and apple during storage by increasing lycopene, total carotenoid, and phenolic compounds content (Pataro et al. [2015\)](#page-19-0) without effects on pH or titratable acidity. UV-C as an abiotic stress induces synthesis of secondary metabolites with antioxidant activities by increasing activities of enzymes in the phenylpropanoid pathway (Cisneros-Zevallos, [2003\)](#page-16-0).

UV-C can also delay the ripening process of fruit. After treated by UV-C, the tomato fruit showed a reduction in respiration rate, ethylene production, and an increase in putrescine (Maharaj, Arul, and Nadeau [1999](#page-18-0)). In addition, it reduced chilling injury of pepper (Vicente et al. [2005](#page-21-0)). However, the impact of UV is related to its dose, treatment time, storage temperature, produce type, and packaging. The various effects of UV-C on quality attributes of fresh and fresh-cut produce are listed in [Table 2](#page-5-0).

Pulsed light as an emerging physical non-thermal technology, is capable of inactivating various microorganisms within a very short time, on food contact surfaces, equip-ment, and food packaging materials (Gómez et al. [2012,](#page-17-0) Ramos-Villarroel et al. [2012a](#page-20-0); Charles et al. [2013\)](#page-16-0). Pulsed light involves the use of short-duration, high-peak pulses of broad-spectrum light (100–1100 nm; UV to near infrared) (Oms-Oliu, Martín-Belloso, and Soliva-Fortuny [2010](#page-19-0)). As a physical preservation method, pulsed light has a positive consumer image. This technology was approved by the U.S. FDA for food processing. It is generally believed that the UV-C component of pulsed light is the most important wavelength region for its bactericidal effects due to the indispensable effect of the UV portion (Ramos-Villarroel et al. [2012a\)](#page-20-0). However, pulsed light is different from UV-C light, because pulsed light destructs microorganisms via multiple mechanisms including inter-related photochemical, photothermal, and photophysical effects (Farrell et al. [2011;](#page-17-0) Kramer and Muranyi [2014](#page-18-0); Rowan, Valdramidis, and Gómez-López [2015\)](#page-20-0). As a multi-target technology, pulsed light is more effective in inactivating microorganisms in a relatively short period of time than continuous UV-light (Scott et al. [2017\)](#page-20-0).

Pulsed light as a relatively novel technology has been evaluated for a number of fresh and fresh-cut produce and exhibits similar beneficial effects as UV-C on fresh produce such as extension of shelf-life, reductions of microbial populations, inhibition of ripening process, and induction of antioxidant compounds (Abida, Rayees, and Masoodi [2014\)](#page-15-0) ([Table 3\)](#page-7-0).

Pulsed light extended the shelf-life of fresh-cut mushrooms with slight effects on texture and antioxidant capacity (Oms-Oliu, Aguilo-Aguayo, et al. [2010\)](#page-19-0). Repetitive pulsed light treatments were effective in extending shelf-life of fresh-cut cantaloupe by maintaining its physical quality (firmness, fluid loss, and color), chemical quality (pH, titratable acidity, total soluble solids, phenolic content and ascorbic acid content), and had minimal effect on tissue structure (Koh et al. [2016\)](#page-18-0). The shelf life of fresh-cut cantaloupes was extended by 20 days at 4° C compared to control in terms of microbiological quality. The extension of shelf-life is due to the reduction of spoilage microorganisms and maintenance of quality attributes. For example, pulsed light reduced populations of native mesophilic bacteria and inoculated Bacillus cereus by 1.0–1.3 log and 1.3–2.0 log, respectively, without causing significant changes on vitamin C, total phenolics, antioxidant capacity, firmness and color of plums, tomatoes, cauliflowers, sweet peppers, and strawberries (Luksiene et al. [2012\)](#page-18-0). Exposure of fresh-cut avocado to pulsed light $(14 \text{ J} \cdot \text{cm}^{-2})$ resulted in reductions of mesophilic microorganisms (1.20 \log (CFU \bullet g⁻¹), delay of the proliferation in yeasts and molds count of fresh cut avocado, and prolonged microbiological shelf life up to 15 days at 4° C (Aguilo-Aguayo et al. [2014\)](#page-15-0). Similarly, intense light pulse (15 or 30 pulses and 0.4 J ecm^{-2} per pulse) decreased ethylene production while inactivating L. innocua and E. coli of avocado cylinders (Ramos-Villarroel, Martín-Belloso, and Soliva-Fortuny [2011](#page-20-0)).

Like UV treatment, pulsed light can induce synthesis of many health-beneficial phytochemicals. For example, pulsed light was useful for fig fruits to enhance red coloration and increase anthocyanin synthesis (Rodov, Vinokur, and Horev [2012\)](#page-20-0). It seemed that pulsed light activated antioxidant defense mechanisms as evidenced by increased activities of superoxide dismutase (SOD) and catalase, and contents of carotenoid, vitamin C, flavonoid, anthocyanin and total phenolics in 'Tommy Atkins' mangoes (Lopes et al. [2016](#page-18-0)). For immature green tomatoes, pulsed light increased lycopene, total carotenoid, phenolic compounds, and antioxidant cap-acity (Pataro et al. [2015\)](#page-19-0). A 5.0 J ecm^{-2} (30 s) pulsed light treatment reduced Salmonella and E. coli O157:H7 populations and improved nutritional quality of raspberries (Xu and Wu [2016\)](#page-21-0).

As with any technology, high doses of pulsed light can result in detrimental effects on sensory and nutritional quality of fresh produce. For example, high doses (12 and 28 J \bullet cm⁻²) had adverse effects on phenolic compounds, vitamin C, and antioxidant capacity of fresh cut mushroom, probably due to thermal damage induced by the treatments (Oms-Oliu, Aguilo-Aguayo, et al. [2010\)](#page-19-0). Pulsed light at doses above $17.5 \text{ kJ} \cdot \text{m}^{-2}$ caused dehydration, browning, and

Table 3. Effects of pulsed light on quality attributes and microbial populations of fresh and fresh-cut produce.

Treatment conditions (concentration, time, intensity, dose etc.)	Type of fresh produce	Effect on quality	Reference
4.8 Jocm ^{-2} 12 and 28 Jocm ⁻² 15days at 4 °C	fresh-cut mushroom	4.8 Jocm ⁻² extended the shelf-life with slight effect on texture and antioxidant capacity; 12 and 28 Jocm ⁻² had adverse effects on phenolic compounds,	Oms-Oliu, Aguiló-Aguayo, et al. 2010
17.5, 52.5, 105.0 and 157.5 kJom ⁻²	apple	vitamin C and antioxidant capacity Doses beyond 17.5 kJ•m ⁻² caused dehydration, browning and	Ignat et al. 2014
15 or 30 pulses and 0.4 Jocm 2 per pulse	avocado	flavor depletion decreased ethylene production and disactivated the L. innocua and E. coli effectively, but the use of 30 pulse caused browning and softening	Ramos-Villarroel et al. 2011
2.7, 7.8, 11.7 and 15.6 J•cm ⁻²	cantaloupe	extended shelf-life with no effect on its physical quality (firmness, fluid loss, color) and chemical quality (pH, titratable acidity, total soluble solids, phenolic content and ascorbic acid content) and minimal effect on tissue structure (7.8 Jocm ⁻² is the best)	Koh et al. 2016
10, 30, 90 and 300 s, with fluence values of 2, 6, 18 and 60 $\text{kl} \cdot \text{m}^{-2}$, respectively	fiq	enhanced red coloration and increased anthocyanin content	Rodov et al. 2012
8 Jocm $^{-2}$	'Kent' mango	maintained firmness, color, carotenoid content, phenol and	Charles et al. 2013
0.6 J cm^{-2}	'Tommy Atkins' mango	total ascorbic acid content activated antioxidant defense mechanisms in pulp (high activity of SOD and CAT, more carotenoid, vitamin C, flavonoid, anthocyanin and total phenolics) and peel (SOD and carotenoid, mangiferin, and total phenolics)	Lopes et al. 2016
1, 2, 4, and 8 Jocm ⁻² each side (0.95, 1.9, 3.8 and 7.6 s each side, respectively) and 2, 4 Jocm ⁻² each side (1 h, 2 h respectively)	green tomato	increased lycopene, total carotenoid, phenolic compounds, and antioxidant capacity	Pataro et al. 2015
3.7 kJom ⁻² UV vs 4.6 kJom ⁻² /pulse (8, 16, 24 pulse)	mature green tomato	high intensity pulsed polychromatic light delayed ripening and enhanced disease resistance	Scott et al. 2017
5.0 Jocm ⁻² 30 s	raspberry	reduced Salmonella and E. coli 0157:H7 and improved nutritional quality	Xu and Wu 2016
2, 10, 20 and 40 s, corresponding to fluences of 2.4, 11.9, 23.9 and 47.8 Jocm ⁻² , respectively	strawberry	reduced softness and fungal incidence	Duarte-Molina et al. 2016
Pulsed UV light dose 5.4 J•cm ⁻² , 5 Hz	plums, tomato, cauliflower, sweet pepper and strawberry	reduced naturally distributed mesophilic bacteria and inoculated Bacillus cereus by 1.0-1.3 log and 1.3-2.0 log, respectively, and no significant change on vitamin C, total phenolics, antioxidant capacity, firmness, and color	Luksiene et al. 2012
3.6, 6.0 and 14 Jocm ⁻² per side	fresh-cut avocado	highest reductions in aerobic mesophilic microorganisms $(1.20 \text{ log } CFU \bullet g^{-1})$ and better hue values and 1.3-fold of chlorophyll a and b were observed after 6.0 Jocm ⁻² treatments.	Aguiló-Aguayo et al. 2014
Intense pulsed light 0, 2, 5, 10, 15, 20 and 30 pulses corresponding to 0, 8, 20, 40, 60, 80 and 120 kJ•m ⁻² , respectively	spinach	a higher respiration rate and antioxidant content just after treatment followed by accelerated deterioration in physicochemical quality	Agüero et al. 2016
1.0 to 3.0 kV (0.1 and 1.0 J•cm ⁻² fluency at a distance of 10 cm)	endive salad and mung bean sprout	a discoloration and high respiration rate of endive salad while an improvement on color without affecting the respiration of mung	Kramer et al. 2015
Full spectrum-light pulses (6 and 12 J•cm ⁻²)	fresh-cut watermelon	bean sprouts negative effects on the color and texture	Ramos-Villarroel et al. 2012b

Table 3. Continued.

flavor depletion of fresh cut apple (Ignat et al. [2014](#page-17-0)). Similarly, pulsed light (30 pulse, 0.4 J ecm^{-2} per pulse) caused browning and softening of avocado cylinders (Ramos-Villarroel, Martín-Belloso, and Soliva-Fortuny [2011\)](#page-20-0). Furthermore, color and texture of raspberries treated with 30 s pulsed light $(28.2 \text{ J} \bullet \text{ cm}^{-2})$ changed negatively during 10 days of storage, while the initial reductions of Salmonella and E. coli could not be maintained (Xu and Wu [2016\)](#page-21-0). Some research showed that intense light pulse might not be applicable on fresh-cut vegetables, such as cabbage and lettuce due to detrimental changes in sensory quality (Gómez-López et al. [2005\)](#page-17-0). In addition, it led to a higher respiration rate after treatment, followed by accelerated deterioration in physicochemical quality of spinach during storage (Agüero et al. [2016\)](#page-15-0), resulted in a discoloration and high respiration rate of endive salad (Kramer, Wunderlich, and Muranyi [2015](#page-18-0)), and had negative effects on the color

and texture for fresh-cut watermelon (Ramos-Villarroel et al. [2012b\)](#page-20-0).

Pulsed light has been combined with coating and surfactants to enhance its effects. For fresh-cut apples, gellan-gum based coating (0.5% w/v) with apple fiber followed by pulsed light (12) \bullet cm⁻²) reduced softness and browning without affecting their flavor (Moreira et al. [2015](#page-19-0)). Furthermore, when combined with coating with alginate, shelf life of fresh-cut cantaloupes was further extended by 28 days compared with samples with the pulsed light alone (Koh et al. [2017\)](#page-18-0). Similar results could be found when pulsed light was combined with pectin-based edible coating, although offodors in fresh-cut apple were detected (Moreira et al. [2017\)](#page-19-0). When berries in water were treated with pulsed light, the efficacy of pulsed light against human pathogen was significantly higher than direct pulsed light (without water). The water-assisted pulsed light treatment had minimal or no

effect on shelf-life, quality attributes, or nutritional compounds (Cao, Huang, and Chen [2017\)](#page-16-0). When combined with quality-stabilizing dip (1% w/v N-acetylcysteine and 0.5% w/v CaCl₂), pulsed light (8 and $16 \text{J} \bullet \text{cm}^{-2}$) helped maintain a better quality and antioxidant characteristics of fresh-cut apples for 15 days at 5° C (Llano et al. [2016\)](#page-18-0). When combined with surfactants such as sodium dodecyl sulfate, pulsed light enhanced microbial safety of green onions with no effect on quality (Xu, Chen, and Wu [2015](#page-21-0)).

Both UV-C and pulsed light are shown to reduce populations of pathogenic and spoilage microorganisms on fresh and fresh-cut produce. In addition, they are capable of increasing the formation of many health beneficial compounds and antioxidant activity. The treatments may induce or inhibit browning of cut surface of fresh produce depending on dose, treatment temperatures, and storage. The treatments, especially pulsed light may result in other undesirable changes on some fresh-cut produce items such as fresh-cut leafy greens. In general, pulsed light is more effective than UV-C because of its high intensity and multiple target mechanisms. Perhaps, the major challenge for the application of UV-C and pulsed light on fresh and fresh-cut produce to enhance microbial safety, is their low penetration ability. The shading effect due to overlap or shading of fresh and fresh-cut produce items prevents uniform and complete light exposure of all produce surfaces. Double sided light treatments or introduction of a rotating/ tumbling device may help to achieve the goal of uniform and complete exposure of UV or pulsed light (Fan, Huang, and Chen [2017](#page-16-0)). To assess UV/pulsed light dose received by a piece of fresh produce, film dosimetry systems may be applied (Yan et al. [2017](#page-21-0)).

Ultrasound

The inactivation of microorganisms by ultrasound is a consequence of cavitation (Majid, Nayik, and Nanda [2015](#page-18-0); Serna-Galvis et al. [2016](#page-20-0)). As ultrasound waves propagate in water or other liquids, small bubbles form. The bubbles grow until reaching a critical size and then collapse violently, causing thermal, mechanical, and chemical effects. de São José et al. ([2014,](#page-16-0) 2015) summarized ultrasound application in fresh fruit and vegetable products including effects on microbiota contamination, enzymes, and food components.

Ultrasound has been investigated in combination with other sanitizers, such as chlorine to enhance their antimicrobial efficacy. Its efficacy is, however, limited, often achieving 1–2 log or less reductions of pathogens on fresh produce after a few minutes of treatment under laboratory settings (de São José et al. [2014](#page-16-0)). For example, Zhou, Feng, and Luo [\(2009\)](#page-21-0) observed that use of ultrasound (200 WL¹) contributed to a reduction of 0.7-1.1 log $(CFU\bullet g^{-1})$ of E. coli O157:H7 in spinach compared with treatments using only chemical sanitizers. Ultrasound in combination with electrolyzed water resulted in 1.77 and 1.29 log reductions on total aerobic bacteria and 1.50 and 1.29 log reductions on yeasts and molds, respectively, for cherry tomatoes and strawberries (Ding et al. [2015](#page-16-0)).

Ultrasound treatment $(106.19 \,\text{W}\bullet \text{L}^{-1}, 25 \,^{\circ}\text{C})$ of cherry tomato fruit in water reduced spoilage microorganisms, delayed the ripening process by inhibiting ethylene production and respiration rate, and maintained color, flavor, firmness, and antioxidant capacity of the fruit (Wang et al. [2015](#page-21-0)). A recent study suggested that bananas that were treated by ultrasound (40 kHz) in water at 25° C for 10 min developed less severity of chilling injury during storage at 5 C (Khademi, Ashtari, and Razavi [2019](#page-18-0)). The combination of ultrasound (25 kHZ, 2 kW, 1 min) with chlorine or peroxyacetic acid did not cause tissue damage measured with electrolyte leakage, total color difference, or firmness during a 14-day storage at 4° C (Salgado et al. [2014\)](#page-20-0). Similar results were reported that the combination of ultrasound (130 W, 42 kHz, 0–30 min) and chlorine did not affect visual quality of fresh-cut lettuce during an 8-day storage at 5 °C (Irazoqui et al. [2019](#page-17-0)). However, confocal imaging indicated that ultrasound damaged the lettuce tissues, resulting in higher microbial growth that occurred during storage in treated samples.

Ultrasound can cause damages to fresh and fresh-cut produce. For example, ultrasound (40 kHz, 200 W for 5 and 10 min) resulted in lower pH and hue angle of fresh-cut potatoes during post-treatment storage (12 °C) (Amaral et al. [2015](#page-15-0)). Combination of ultrasonic treatment and electrolyzed water reduced firmness of cherry tomatoes, while other quality attributes such as soluble solids and acidity were not affected (Ding et al. [2015](#page-16-0)). Mustapha et al. [\(2020](#page-19-0)) recently reported that ultrasound (10 min, 20/40 kHz) alone or in combination with peracetic acid (40 mg \bullet L⁻¹) or H₂O₂ (5%) resulted in loss of tomato firmness, even though up to 3.1 log reductions of natural microbiota were observed. Another study showed that lettuce leaves treated with ultrasound (26 kHz, 200 W 5 min) had lower scores of sensory quality attributes after 10 days of storage at 5° C (Neto et al. [2019](#page-19-0)). After Romaine lettuce leaves were treated with ultrasound (25 kHz) at 26 W/L for 1–3 min, quality was not immediately affected (Yu, Engeseth, and Feng [2016](#page-21-0)). During 60 h storage at ambient temperature, an increase in phenylalanine ammonia lyase activity was observed, resulting in formation of phenolic compounds and enhancement of antioxidant capacity. The changes in quality of fresh produce due to ultrasound are related to the burst of cavitation bubbles, which results in localized high temperature and pressure. The transient high pressure and temperature cause the rupture of the cell membrane leading to the cellular leakage, loss of turgor pressure and tissue softening. The loss of tissue integrity also results in changes in enzymatic reactions and consequently the formation of secondary metabolites.

In summary, ultrasound-assisted washing of fresh produce has shown to enhance sanitation efficacy of aqueous sanitizers mainly due to the physical effects of acoustic cavitation (Palma, Zhou, and Feng [2017\)](#page-19-0). However, its efficacy is often limited achieving \sim 1 log reduction compared with sanitizers themselves. In addition, cavitation and burst of bubbles during ultrasound treatment would also disrupt cell

Table 4. Effects of high pressure on quality attributes and microbial populations of fresh and fresh-cut produce.

Treatment conditions (concentration, time,			
intensity, dose etc.)	Type of fresh produce	Effect on quality	Reference
400 MPa for 2 min at 4 °C	iceberg lettuce and strawberry	inactivated murine norovirus with a slight influence on food quality	Lou et al. 2011
High pressure carbon dioxide treatment (12 MPa, 40 °C, 15 min)	fresh-cut carrot	maintained bioactive compound (phenol, flavonoid, carotenoid, antioxidant capacity) and enzyme stability and inactivated the natural microbial flora though texture and ascorbic acid loss	Spilimbergo et al. 2013
High pressure argon treatment (150 MPa for 10 min)	fresh-cut apple	delayed browning and microbial growth, reduced total phenolics loss without effect on titratable acidity and soluble solids for at cold storage	Wu, Zhang, and Wang 2012
High pressure argon treatment $(1.8 \text{ MPa}, 60 \text{ min } 4 \degree \text{C})$	fresh-cut pineapple	extended shelf-life	Wu, Zhang, and Adhikari 2012
High pressure nitrogen treatment (120 MPa 10min)	fresh-cut pear	inhibited microbial growth, maintained sensory quality, and improved total phenolics without affecting titratable acidity and soluble solids	Xu, Liu, et al. 2015
high-pressure carbon dioxide and high- power ultrasound (12 MPa, 10 W, 40 °C, 20 min)	fresh-cut coconut	8 log reductions of S. Typhimurium with almost no effect on physicochemical quality including total acidity, pH, texture, color, enzymatic activity, antioxidant capacity, flavonoids, phenolic acids, phenols and dry matter, fat content	Ferrentino et al. 2015
peroxyacetic acid (100 mg $\bullet L^{-1}$) dip for 2min in combination with: (a) vacuum (V: 10 mbar) or (b) positive pressure application $(P: 3 bar)$.	lettuce	reduced microorganism population, it also had detrimental effect on the visual quality	Petri et al. 2015
400 MPa and 5 min for HHP, 1.2×10^5 Jom ⁻² for pulsed light, modified chitosan containing a nanoemulsion of mandarin essential oil	green beans	had antagonistic effect against L. innocua though with impact on green beans color and firmness	Donsì et al. 2015

wall and membrane structures of fresh produce causing cellular leakage, resulting in the loss of turgor pressure and consequent changes in texture, especially to fresh-cut leafy greens. Because of electric leakage, compromised tissue integrity, and consequent availability of nutrients to microorganism, the initial reduction of microorganisms may not be maintained during post-treatment storage. Taken together, the commercial application of ultrasound to enhance microbial safety of fresh and fresh-cut produce remains a challenge.

High pressure processing

High hydrostatic pressure (HHP) processing, also called high pressure processing, uses hydrostatic pressures commonly in the range of 100–800 MPa to treat various food products (Martínez-Monteagudo and Balasubramaniam [2016\)](#page-18-0). Application of the high pressure for a period of 30 s to a few minutes inactivates microorganisms including pathogenic bacteria, native microflora, yeast and mold in foods, therefore enhancing microbial safety and extending shelf life (Hwang and Fan [2015;](#page-17-0) Wang et al. [2016](#page-21-0)). The inactivation of microorganisms is due to pressure induced damage on cell membrane and cellular integrity, which result in altered cell permeability, loss of osmotic regulation and genetic functions, and altered biochemical reactions (Barba et al. [2015\)](#page-15-0) (Table 4).

While high pressure induces cellular injury to microorganisms, it certainly can cause similar damage to plant cells as well. For example, high pressure (300 MPa) led to a

significant detrimental effect on the visual quality of lettuce (Petri, Rodríguez, and García [2015\)](#page-19-0). High pressure (400–600 MPa for 2 min at 4° C) resulted in considerable textural loss of blueberries, strawberries, and raspberries due to pressure-induced softening of the tissue even though the treatment inactivated murine norovirus, a surrogate for human norovirus, in low temperature and neutral pH (Lou et al. [2011\)](#page-18-0). HHP (400 MPa 5 min) with or without combinations of chitosan coating or pulsed light had a significant impact on firmness and color of green beans even though HHP had antagonistic effects on Listeria innocua (Donsì et al. [2015](#page-16-0)). HHP treatment disturbs the cell permeability of fruits and vegetables. This altered cell permeability enables movement of water and metabolites in the cell. The degree of cell disruption is not only dependent on the extent of applied pressure level but also on the type of plant cell (Oey et al. [2008\)](#page-19-0).

Because high pressure caused significant changes in the quality of fresh produce, especially in texture, relative lower pressure has been investigated often in combination with pressurized gases. High pressure carbon dioxide treatment (12 MPa, 40 °C, 15 min) (pressured $CO₂$) of fresh-cut carrots maintained bioactive compounds (phenol, flavonoid, carotenoid, antioxidant capacity) and microbial stability for 4 weeks at 4° C. However, texture and ascorbic acid losses were 90% and 40% respectively (Spilimbergo et al. [2013\)](#page-20-0). The high pressure treatment at a lower temperature $(22^{\circ}C)$ did not exhibit microbial stability after 2 weeks of storage, due to a faster growth rate during storage compared with untreated samples.

High pressure nitrogen treatment (120 MPa 10 min) of fresh-cut pears inhibited microbial growth, maintained sensory quality, and improved total phenolics without affecting titratable acidity and soluble solids during storage (Xu, Liu, et al. [2015](#page-21-0)). High pressure argon treatment (150 MPa for 10 min, 4° C) delayed browning and microbial growth, and reduced total phenolics loss without effects on titratable acidity and soluble solids for fresh-cut apples during storage at 4° C (Wu, Zhang, and Wang [2012\)](#page-21-0). Application of high pressure argon treatments maintained quality of fresh-cut pineapples during cold storage and extended the shelf life by 6 days at 4° C (Wu, Zhang, and Adhikari [2012\)](#page-21-0). The delay in the growth of microbes was probably due to argon or nitrogen dissolved in pineapple tissues and reduced water activity.

Combination of high-pressure carbon dioxide and highpower ultrasound technique (12 MPa, 40 °C, 20 min) achieved 8 log reductions of S. Typhimurium with almost no effect on physicochemical quality including total acidity, pH, texture, color, enzymatic activity, antioxidant capacity, flavonoids, phenolic acids, phenols and dry matter, or fat content of fresh-cut coconut (Ferrentino, Komes, and Spilimbergo [2015](#page-17-0)).

In general, to achieve significant reductions $(>1 \log)$ of pathogenic bacteria at ambient or low temperatures, the pressure of 300-600 MPa was required. For example, HHP (300 MPa, 8 C for 5 min) inactivated populations of Salmonella and Listeria spp. in cantaloupe puree by 2.4 ± 0.2 and $1.6 \pm 0.5 \log (CFU \bullet g^{-1})$, respectively (Mukhopadhyay et al. [2016\)](#page-19-0). The pressure at the range (300-600 MPa) likely results in adverse changes in the quality of fresh and freshcut produce. As mentioned earlier, pressure affects cell wall and membrane, which are associated to cell turgor of fresh and fresh-cut produce. At low pressure levels, the effect of cell turgor is reversible, reflected by the reestablished membrane semi-permeability and functionality of membranebound proteins. However, above a certain threshold, the effect of pressure become irreversible, resulting in tissue damage (Rux et al. [2019,](#page-20-0) [2020\)](#page-20-0). The threshold was 100 MPa for most fresh and fresh-cut produce. One of the major effects of HHP is loss of firmness due to the failure of maintaining cell turgor and the overall tissue structure of the produce.

Fresh and fresh-cut produce are 'living' organisms and tissues. A living organism or tissue undergoes physiological processes such as respiration and other controlled metabolism activities. Furthermore, it can respond to abiotic and biotic stresses by synthesizing defense related compounds, such as secondary metabolic compounds (Toivonen and DeEll [2002](#page-20-0)). After treatments with high pressure at pressure above the thresholds, the fresh or fresh-cut produce can probably not be regarded as 'living' organisms because the cell integrity is lost and tissues fail to orchestrate a response to stresses, even though certain activities such as enzymes may be active.

Another characteristic of a living plant or tissue is compartmentation of metabolism and other cellular functions (Lunn [2007\)](#page-18-0). The loss of cellular compartmentation due to HHP will allow release of enzymes and other compounds which come in contact with one another, resulting in dysfunction of metabolism. Therefore, it is questionable that HHP can be used to treat fresh and fresh-cut produce to enhance microbial safety of fresh produce, although combination of HHP with other techniques, such as pressurized gases may allow the use of low pressure to minimize the damage to fresh produce. It is also debatable whether the products treated by HHP can be claimed as fresh or freshcut produce, while they can certainly be regarded as minimally processed as HHP maintains a better sensory and nutritional quality than traditional thermal treatment (such as canning processing). Due to the cost associated with HHP, its use may be only for niche market and high value products.

Cold plasma

Plasma refers to ionized gas consisting of particles including photons, free electrons, excited or non-excited atoms, and positive or negative ions and molecules (Fernández et al. [2012](#page-17-0)). In addition, UV light and oxidants such as ozone and hydrogen peroxide are produced during the processing. Cold plasma is also referred as nonthermal plasma or atmospheric cold plasma because it is produced at ambient temperature under normal atmospheric pressure. It has been demonstrated that cold plasma inactivates pathogenic and spoilage microorganisms on various foods and surfaces (Bourke et al. [2018\)](#page-16-0). Factors affecting the efficacy of cold plasma include generating device, power input, mode of exposure, exposure time, feeding gas or atmosphere composition, temperature, and relative humidity. Due to the nature and versatility of cold plasma, the mechanisms for its effects on fresh produce are rather complex. Its impact is related to cold plasma-induced reactive species, change in pH, UV light, formation of oxidative compounds $(O_3, H_2O_2$ etc.) and the interactions of all above entities.

Cold plasma has been reviewed as a novel decontamination technology for fresh and fresh-cut produce (Niemira [2012](#page-19-0); Mir, Shah, and Mir [2016;](#page-19-0) Misra [2016](#page-19-0); Bourke et al. [2018](#page-16-0); Pankaj and Keener [2018](#page-19-0); Pankaj, Wan, and Keener [2018](#page-19-0)). [Table 5](#page-12-0) summarizes recent studies on cold plasma effects on quality of fresh and fresh-cut produce. Studies have demonstrated that cold plasma has no or limited negative impact on sensory or nutritional quality attributes of fresh and fresh-cut produce while reducing populations of pathogenic and spoilage microorganisms and extending shelf-life of fresh and fresh-cut produce.

Cold plasma can be used to treat fresh and fresh-cut produce inside sealed packages (Misra et al. [2019](#page-19-0)). For example, an in-package dielectric-barrier discharge (DBD) system (called PlasmaLabel) was able to produce more than 1,000 ppm of ozone inside sealed package containing cherry tomatoes within 1 min of treatment (Fan et al. [2012\)](#page-17-0). The system achieved more than 4 log reductions of L. innocua and 2–3 logs reductions of Salmonella and E. coli. No negative effects on tomato color or texture were observed during a 22-day post-treatment storage at 10° C. Similarly, in-

Table 5. Effects of Cold plasma on quality attributes and microbial populations of fresh and fresh-cut produce.

Table 5. Continued.

Treatment conditions (concentration,			
time, intensity, dose etc.)	Type of fresh produce	Effect on quality	Reference
20 and 40 min (for 2 sides)	kiwifruit	maintained color, texture, and antioxidant activity	Ramazzina et al. 2015
treated for 45 s with hydrogen peroxide (7.8%) aerosols activated by atmospheric cold plasma followed by additional 30 min dwell time	grape tomato, baby spinach and cantaloupe	inactivated human pathogens by up to 4.9 log CFU/piece with slight effect on color and texture	Jiang et al. 2017
60 s at a fixed power (8 W) and 5 Lomin ⁻¹ of argon mixed with 0.1% oxygen	corn salad, cucumber, apple, and tomato	reduced E. coli DSM 1116 by up to 4.7 ± 0.4 log	Baier et al. 2014
Plasma processed air treatment for 2.5, 5, 10min	apple, carrot, tomato and cucumber	10 min treatment reduced inoculated E. coli by 4.6 ± 2.0 and 6.0 ± 0.8 log cycles on apples and carrots, respectively, however, significant effects were found on color of tomatoes and carrots, and on chlorophyll fluorescence parameters of cucumbers	Baier et al. 2015
clove oil (1 mg \bullet mL ⁻¹ , 2 mg \bullet mL ⁻¹ and $4 \text{ mg} \cdot \text{mL}^{-1}$ 30 min) and cold nitrogen plasma (400 W 3min for 2 sides)	lettuce	synergetic antibacterial efficacy against E. coli 0157:H7 biofilms but with mild effect on sensory quality	Cui et al. 2016
non-thermal plasma-activated water for 0.5, 2, or 5 min	Chinese bayberry	0.5min achieved a maximum reduction of 1.1 log CFU/g both for bacteria and fungi and reduced decay and improved firmness, color and total soluble solids	Ma et al. 2016.
Atmospheric air DBD (Dielectric barrier discharge) plasma discharge system (36V, 1.8A) plasma treatment for 0, 2, 4, 6, 8 and 10 min	blueberry	Populations of bacteria and fungi decreased by 93.0% and 25.8%, respectively in the 10 min group, and the blueberry decay rates reduced by up to 17.7%. Contents of sugar, vitamin C, and total anthocyanin as well as the superoxide dismutase activity level showed the maximum increases of 1.5-fold, 1.5-fold, 2.2-fold, and 79.3% in the plasma treatment groups of 6, 8, and 10 min, respectively, after 20 days of storage.	Dong and Yang 2019
dielectric barrier discharge plasma at 45 kV for 1 min	strawberry	promoted the accumulation of total phenolics, total flavonoid, anthocyanin, maintained the texture properties and inhibited microbial growth	Li et al. 2019
60, 80, and 100 kV (60 Hz frequency) for 1, 2, 3, 4, and 5 min	fresh-cut carrot	100 kV for 5 min reduced about 2 log in total aerobic mesophiles, and yeast and mold, minor changes in pH, color, texture, and total carotenoids.	Mahnot et al. 2020

package treatment of cherry tomatoes with high-voltage DBD system for 50 s achieved greater than 5 log reductions of E. coli populations on tomatoes and a maximum of 3.5 log unit reduction for *L. innocua* with no significant difference in color, firmness, pH, or total soluble solids observed between control and treated samples (Ziuzina et al. [2016\)](#page-21-0). The treatment produced 450–900 ppm of ozone inside the packages. Furthermore, the in-package DBD treatment system was effective against L. innocua inoculated on strawberries, with 3.8 log reductions achieved. No significant impact on color, firmness, pH or total soluble solids was observed (Ziuzina et al. [2020\)](#page-21-0). However, the in-package treatment slightly degraded ascorbic acid and anthocyanins content of strawberry (Misra et al. [2015\)](#page-19-0).

The reductions of pathogenic bacteria were frequently less on fresh-cut produce than on whole fruits with smooth surface. For example, cold plasma treatment reduced populations of inoculated E. coli O157:H7 and Salmonella Typhimurium on fresh-cut lettuce by up to 2.8 log $(CFU•g^{-1})$ without loss of physicochemical or sensory properties (Song et al. [2015\)](#page-20-0). Dielectric barrier atmospheric cold plasma (DACP) reduced the initial counts of E. coli O157:H7 and total aerobic microorganisms by only

approximately 1 \log (CFU \bullet g⁻¹) without significantly affecting the color, $CO₂$ generation, weight, and surface morphology of fresh-cut Romaine lettuce during storage (Min, Roh, Boyd, et al. [2017](#page-18-0)). The reductions of E. coli O157:H7 populations on romaine lettuce leaves with a 5-layer configuration in a package were only 0.4–0.8 log CFU per piece of lettuce without significant changes on the surface morphology, color, respiration rate, or weight loss of the samples (Min, Roh, Niemira, et al. [2017\)](#page-18-0).

Although fresh and fresh-cut produce treated with cold plasma in general maintains its quality during post-treatment storage, too long or severe treatment of fresh produce with cold plasma can cause damages to fresh produce. For example, cold plasma significantly reduced anthocyanin content, firmness, and color parameters $(L^*$ and a^{*} values) of blueberries after 90, 60, and 120 s treatments, respectively (Lacombe et al. [2015\)](#page-18-0). However, another study (Dong and Yang [2019](#page-16-0)) showed that DBD cold plasma increased anthocyanin content of blueberry by up to 79.3%, possibly due to increased extraction efficiency of anthocyanin.

It has been shown that cold plasma impacts some enzyme activities. For example, polyphenol oxidase activity was reduced by about 62% and 77% in fresh cut apples and

	Pathogen			Challenges for
Technology	inactivation mechanism	Penetration ability	Potential quality deterioration	commercialization
lonizing irradiation	Radicals and reactive species from radiolysis of water	High (gamma and X-rays)	Softening, vitamin C loss	Consumer acceptance
UV/pulsed light	DNA damage	Surface	Surface browning	Limited penetration
Ultrasound	Membrane damage	Surface	Softening	Prorogation of ultrasound in large scale
High hydrostatic pressure	Cavitation	Penetration	Softening	Tissue damage, cost
Cold plasma	Radicals and reactive species	Surface	Surface discoloration	Limited research, regulatory approval

Table 6. Comparison of non-thermal processing technologies: mechanism, penetration ability, potential quality damages and challenges for commercialization.

potato tissue, respectively, following exposure to plasma processed air for 10 min (Bußler, Ehlbeck, and Schlüter [2017\)](#page-16-0); peroxidase was even less stable and reduced by about 65% and 89% in fresh cut apple and potato tissue, respectively. The pH value on the tissue surface dropped to 1.5 while cell integrity was not significantly affected (Bußler, Ehlbeck, and Schlüter [2017](#page-16-0)). The decrease in pH was probably due to the formation of acidic chemical species such as HNO3, and ONOOH (Pan, Cheng, and Sun [2019](#page-19-0)). The decreased surface pH and reduced activity of the enzymes led to slower tissue browning (Tappi et al. [2014\)](#page-20-0).

Cold plasma can be combined with other antimicrobials to achieve higher reductions of pathogen. Treatments with cold nitrogen plasma and clove oil resulted in synergetic antibacterial efficacy against E. coli O157:H7 biofilms on lettuce. However, the appearance and overall acceptability of the treated lettuce were lower than the nontreated samples (Cui, Ma, and Lin [2016](#page-16-0)). Cold plasma was also used to activate (ionize) hydrogen peroxide aerosol by passing aerosolized hydrogen peroxide through plasma field (Song and Fan [2020\)](#page-20-0). The combined treatment significantly $(P < 0.05)$ improved the efficacy of aerosolized hydrogen peroxide against Salmonella and L. innocua on tomatoes, apples, cantaloupe, and Romaine lettuce. Firmness and color of treated samples (tomato, baby spinach and cantaloupe) were not significantly affected (Jiang et al. [2017](#page-17-0)). Research also showed that cold plasma could enhance antibacterial activity of green tea extract on fresh-cut dragon fruit (Matan et al. [2015\)](#page-18-0).

Plasma activated water, obtained from treatment of water with cold plasma, has been applied as a wash to treat fresh and fresh-cut produce. Reactive species produced from oxygen and nitrogen in air by cold plasma are dissolved in water, and reactive species in water along with low pH have antimicrobial activities against various bacteria (Thirumdas et al. [2018](#page-20-0); Xiang et al. [2019\)](#page-21-0). Washing fresh-cut pears with plasma-activated water for 5 min significantly reduced the growth of native microbiota on fresh-cut pears during a 12 day storage at 4° C, and did not affect soluble solid content and titratable acidity (Chen et al. [2019\)](#page-16-0). However, the presence of organic and inorganic materials leached from freshcut produce may reduce the efficacy of plasma activated water against bacteria when used for commercial application.

Cold plasma is an emerging technology that has a potential to be used to mitigate the risk of pathogen contamination without significant effects on product quality of fresh and fresh-cut produce. However, studies conducted so far have been performed mostly in the lab-scale, where only a few pieces of fresh produce are often treated. When scaled up, the technology will likely have limited effectiveness on human pathogens present on fresh produce. The technology needs to be validated in pilot scale and commercial trials. Some other limitations of cold plasma technology are the extended treatment times and the need for the sample to be in proximity to the plasma source for certain types of cold plasma applications. Despite such limitations, there is a potential for plasma being incorporated into packing line of fresh-cut produce processing facilities. Specifically, cold plasma can be combined with modified atmosphere packaging to maintain the quality of fresh-cut produce during post-treatment storage (Misra, Moiseev, et al. [2014\)](#page-19-0). Super atmospheric oxygen packaging in which packages are flushed with high oxygen may be used to promote the efficacy of cold plasma treatment because high amount of reactive species is produced in presence of high oxygen levels.

Considerations for commercialization and research directions

Fresh and fresh-cut produce provides consumers convenience and health benefits, meeting the changes in lifestyle and the demands for healthy foods. However, outbreaks of foodborne diseases have been associated with these products in recent years. Several non-thermal physical technologies have been developed and evaluated on fresh and fresh-cut produce to minimize the risk of human pathogen contamination. It is obvious that each of the technologies has its own challenges for commercial applications, such as limited antimicrobial efficacy, undesirable changes in product quality, requirement of regulatory approval, and reluctance in consumer acceptance (Table 6). Regarding regulatory approval, all physical interventions discussed in this review except cold plasma have been approved by U.S. regulatory agencies for use on fresh and fresh-cut produce, although there are restrictions in terms of dose, intensity, types of fresh produce, and purposes of use. As outbreaks of foodborne illnesses associated with fresh produce occur every year, it is important to continue research on microbial inactivation technologies, which may eventually be applicable to the produce industry to market safer fresh produce (Julien-Javaux et al. [2019](#page-17-0)). To facilitate the commercialization of the technologies, cost analysis, sustainability, and consumer education should also be considered. For example, ionizing radiation is very effective in reducing populations of fresh produce and has been approved for use on some fresh and fresh-cut produce, however consumers are reluctant to

accept the technology due to misconception of the technology. In addition, some technologies such as cold plasma are still in an early stage of development and only laboratory scaled experiments are conducted. Therefore, optimization and industrial scale trials are required before being considered for commercial trials. In addition, possible formation of chemical byproducts needs to be evaluated for regulatory approval. Moreover, one technology may not be effective enough in inactivating human pathogens without causing changes in various quality attributes. Therefore, combinations of technologies with other treatments may be desirable. Finally, to develop intervention technologies that address the microbial food safety issues associated with fresh and freshcut produce, it is clear that a multidisciplinary strategy is needed that involves microbiology, engineering, food quality, and postharvest biology.

Conclusion

This article reviews recent developments in the technologies with a focus on their impact on quality attributes of fresh and fresh-cut produce. Ionizing radiation at doses of 1 or 2 kGy does not significantly impact the quality of most fresh and fresh-cut produce items. Yet, the commercial application of the technology is very limited partially due to the reluctance in consumer acceptance. UV-C and pulse light exhibit some quality benefits, such as increasing antioxidants. However, owing to their low penetration, it is difficult to significantly reduce populations of pathogens on all surfaces of fresh produce. While ultrasound may enhance the antimicrobial efficacy of aqueous sanitizers, the efficacy is rather limited, often achieving about 1 log reductions of common pathogenic bacteria and causing quality deterioration. While high hydrostatic pressure has been studied on fresh and fresh-cut produce demonstrating that high pressure (>300 MPa) significantly inactivates pathogenic bacteria, though product quality, especially firmness, is compromised. In addition, whether high pressure-treated products can be regarded as fresh produce is debatable. Cold plasma is an emerging technology showing limited effectiveness on quality while achieving significant reductions of human pathogens. Much more research is needed before being considered for commercial application. Overall, there are a number of physical technologies that can enhance microbial safety of fresh and fresh-cut produce. However, each of the technologies has limitations and further research is needed to minimize quality loss, while maximizing antimicrobial efficacy.

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